

RESEARCH

Open Access



Spectrophotometric resolution for quantitative analysis of aspirin and rivaroxaban combination therapy in biological fluids using simple and eco-friendly procedure

Heba M. Mohamed^{1*} and Hebatallah M. Essam¹

Abstract

Patients diagnosed with symptomatic peripheral artery disease (PAD) in the lower extremities have a higher likelihood of suffering from major vascular events. Recently, FDA has approved the combination therapy of aspirin (ASP) and rivaroxaban (ROX) to reduce acute limb ischemia and other comorbidities in (PAD) patients. Zero order and ratio absorption spectra were employed in three simple and accurate spectrophotometric techniques (dual wavelength (DW), ratio difference (RD) and derivative ratio (¹DD) for concurrent detection and quantification of ASP and ROX in their pure forms, lab synthetic mixtures and in biological fluid. Our approach involves careful parameter optimization, including solvent selection, sample volumes, and instrumental settings, to reduce the analysis environmental impact. The acquired recovery percentages of accuracy were within 98–102% for pure active pharmaceutical ingredients and 90–110% for pharmaceutical formulations and biological determinations. A comprehensive assessment was done to compare the three methods regarding their ease of use, linearity, sensitivity, conditions, and limitations. The specificity of the proposed methods was evaluated by analyzing the lab synthetic mixtures. The suggested spectrophotometric methods were validated in compliance with ICH guidelines to confirm the validity claims. Also, statistical analysis was done to compare the outcomes obtained from the suggested methods with those obtained from the official ones and they agreed with null hypothesis regarding accuracy and precision. Furthermore, a comprehensive assessment of the environmental sustainability of the developed method was carried out using the Analytical Greenness Calculator, AGREE algorithm. The selected drugs can be efficiently, safely and economically analyzed by the suggested methods in pharmaceutical and biological matrices with no pretreatment or preliminary separation steps and thereby increasing their greenness level.

Keywords Aspirin, Rivaroxaban, Peripheral artery disease (PAD), Biological fluid, Greenness assessment

Introduction

Peripheral artery disease (PAD) is a serious condition that often goes disregarded or even unaddressed by both patients and healthcare providers due to insufficient awareness and other health issues that take the precedence [1]. Globally, about 200 million individuals are affected by peripheral artery disease (PAD), which is

*Correspondence:

Heba M. Mohamed
heba.moustafa@pharma.cu.edu.eg

¹ Analytical Chemistry Department, Faculty of Pharmacy, Cairo University, Kasr Al-Aini St., Cairo 11562, Egypt



related to the considerable cardiovascular adverse events, morbidity, and mortality [2, 3]. If PAD is timely diagnosed and managed, cardiovascular risks can be highly decreased and maintained [1]. Recently in 2023, Janssen Pharmaceuticals (Johnson & Johnson) has revealed data from phase 3 VOYAGER PAD clinical trial, which supports the benefits of prescribing rivaroxaban in combination with aspirin in the management of disease over the use of aspirin alone (the current standard care) [4]. This combination therapy becomes the first approved one by FDA in April 2021 [5, 6]. In addition, this combination provides 33% reduction in acute limb ischemia following Lower Extremity Revascularization (LER) [5]. Aspirin, [2-(acetyloxy) benzoic acid] or acetylsalicylic acid, is one of the non-steroidal anti-inflammatory drugs (NSAID) that exhibits anti-inflammatory, analgesic, and antipyretic effects. Aspirin, structure shown in Fig. 1, is also widely used for its antithrombotic properties, which result from its ability to reduce platelet aggregation [7]. Rivaroxaban, structure shown in Fig. 1, is an oral anticoagulant, a potent, selective, and direct inhibitor of factor Xa that prevents thromboembolism during surgical operations [8]. Rivaroxaban has demonstrated efficacy in both preventing and treating venous thromboembolism, also, it can be used to prevent stroke in individuals with atrial fibrillation. Furthermore, rivaroxaban is the only anticoagulant approved so far to be taken in combination with aspirin to reduce the risk of major thrombotic vascular events in peripheral artery disease (PAD) patients. It is also worth mentioning that in some cases fatal adverse effects related to rivaroxaban combined with aspirin were reported in some patients which recommends the proper monitoring of coagulation during rivaroxaban treatment along with proper attention to dosage error [9, 10].

Aspirin had been analyzed either in single formulation or in multi-component medications using UV-VIS spectrophotometry [11–16], chromatography; HPLC [17–24], UPLC [25] and TLC methods [26, 27] and electrochemical analysis [28, 29]. While, for rivaroxaban, the literature shows determination of ROX by spectrophotometric methods [30, 31] and many chromatographic methods for quantification of ROX in its dosage form via HPTLC [32, 33], HPLC [34–36] and in plasma [37, 38]. Electrochemical behavior investigation using square-wave voltametric to determine rivaroxaban in pharmaceutical dosage forms was also reported [39].

Since this combination therapy was just approved in March 2023, literature check showed no publications of simple and accurate spectrophotometric methods to analyze this novel therapeutic protocol. Therefore, the aims of this work are, first to develop and validate three smart and simple spectrophotometric methods to resolve severe overlapping spectra of ASP and ROX without previous separation. These methods rely on either using the zero-order spectra (dual wavelength) or the ratio spectra (ratio difference and derivative ratio) to resolve the overlap and allow the determination of aspirin and rivaroxaban in their pure forms and laboratory synthetic mixtures containing their therapeutic regime ratio. Furthermore, the suggested Spectro-photometric methods were successfully applied to determine the drugs in biological fluids (human plasma). The methods are simple, accurate and precise, moreover, they do not require any sophisticated apparatus or complicated software. Besides, minimum use of solvents, energy, and hazardous wastes during the whole analysis identifies the suggested methods as economic and eco-friendly ones.

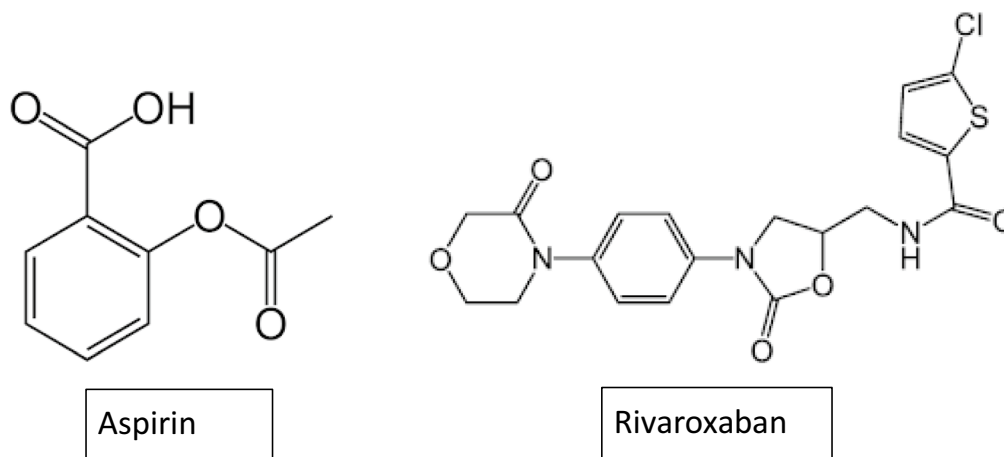


Fig. 1 Chemical structure of Aspirin and Rivaroxaban

Experimental conditions

Apparatus and software

Double beam spectrophotometer: Shimadzu (UV-1900i, Japan) using matched 1.00 cm quartz cells was employed to carry out spectrophotometric measurements. Scans were carried out in the range from 200 to 350 nm at 0.2 nm intervals. Spectra were automatically obtained by Shimadzu UV-Probe 2.62 system software. Centrifuge (Model: 2-16P, Sigma Laborzentrifugen, Germany), PVDF filter (Millex1-GV, Merck Millipore, Cork, Ireland).

Samples

Standards

Aspirin (ASP) was purchased from Sigma laboratories, its purity was tested and found to be $100.22 \pm 0.947\%$ as determined by the official titrimetric USP method [40]. Rivaroxaban (ROX) was kindly supplied from Megafine Pharma (P) Ltd., its purity was tested and found to be $100.48 \pm 0.915\%$ as determined by reported method [34].

Pharmaceutical formulation

Aspirin protect 100 mg[®] by Bayer Schering Pharma, Germany batch # BTAB235, labeled to contain 100 mg ASP/tablet, was purchased from market.

Xarelto[®] by Bayer Schering Pharma, Germany batch # NDC 50458–580-30 claimed to contain 10 mg ROX/tablet, was purchased from market.

Materials and solvents

Methanol and DMSO of HPLC grade, was obtained from Sigma- Aldrich, Darmstadt-Germany.

Blank human plasma samples were obtained from El-Kasr El-aini Hospital Blood Bank and kept frozen until use after gentle thawing.

Standard solutions

Stock standard solution 100 µg/mL of aspirin in methanol and stock standard solution 100 µg/mL rivaroxaban in methanol: DMSO mixture (95:5) were freshly prepared and stored in the fridge during analysis time.

Laboratory synthetic mixtures containing different ratios of ASP and ROX

Into a series of 25-mL volumetric flasks, variable aliquots of ASP and ROX were transferred from their corresponding stock solutions (100 µg/mL). Then volumes were completed with methanol to prepare mixtures containing

different ratios of the two drugs, including the recommended combination therapy ratio.

Procedure

Spectral characteristics

Zero order absorption spectra of ASP and ROX each of concentration 8 µg/mL and a mixture of both components (4 µg/mL each) in methanol were recorded over the range 200–350 nm, as shown in Fig. 2.

Spectrophotometric measurements

Aliquots of 1.0, 2.0, 10.0 mL from ASP stock standard solution (100 µg/mL) in methanol and aliquots of 0.5, 1.0, 2.0, 5.0 mL from ROX stock standard solution (100 µg/mL) in methanol: DMSO mixture (95:5) were accurately transferred into a series of 25-mL volumetric flasks and the final volumes were completed with methanol. Then the absorption spectra of these standard solutions were scanned in the specified wavelength range (200–350 nm) using methanol as a blank and saved in computer to be used for subsequent data manipulation and spectral resolution of the two drugs.

Spectrophotometric methods using zero-order spectra.

Dual wavelength METHOD (DW) Absorbance differences of zero order absorption spectra (⁰D) of the prepared ASP solutions (4.0–40.0 µg/mL) were measured between 242.5 nm and 255.5 nm, where they show significant values, while those of ROX are zeros. On the other hand, for determination of ROX; the absorbance differences of (⁰D) of its prepared solutions (2.0–20.0 µg/mL) were measured between 252 and 286 nm that correspond to ROX concentrations, while those of ASP are having zero values between the selected wavelengths. Formerly the absorbance difference between the two designated wavelengths for each drug was plotted versus the differ-

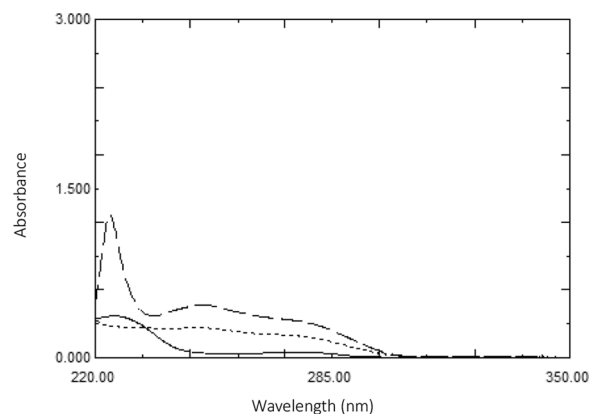


Fig. 2 UV zero order absorption spectra of (8 µg/mL) ASP (—) and ROX (---) and a mixture of 4 µg/mL each (.....), all in methanol

ent ASP and ROX concentration values and the regression equations were computed.

Spectrophotometric methods manipulating ratio spectra

Ratio difference method (RD) The stored spectra of ASP and ROX were used to obtain the ratio spectra. For ASP, its absorption spectra were divided by the 20 $\mu\text{g/mL}$ ROX spectrum, while for ROX, its spectra were divided by 36 $\mu\text{g/mL}$ ASP spectrum. The difference between amplitudes of the ratio spectra at the selected wavelength pairs (234 nm & 280 nm for ASP and 254 & 297 nm for ROX) were plotted against the corresponding concentrations of ASP and ROX, respectively to construct the calibration graphs from which the regression equations were computed.

Derivative ratio method (^1DD) The obtained ratio spectra of ASP & ROX by dividing their zero order spectra by 20 $\mu\text{g/mL}$ ROX spectrum or 36 $\mu\text{g/mL}$ ASP spectrum, respectively, were transformed into the corresponding first order derivative spectra. Derivatization was applied with $\Delta\lambda = 4$ nm that allow minimum noise with more detailed spectral features, and a scaling factor = 10 to achieve suitable measurements. Peak amplitudes at 238.5 nm and 293 nm were measured and plotted against ASP and ROX concentrations respectively, to construct the calibration graphs from which the regression data was obtained for each drug.

Analysis of laboratory synthetic mixtures

Spectrophotometric methods using zero-order spectra

To determine ASP and ROX by DW method, scan the prepared mixtures in the range of 200–350 nm, measure the differences in absorbance between the selected pair of each drug (242.5 nm & 255.5 nm for ASP and 252 nm and 286 nm for ROX). Then the drugs concentrations were calculated using the corresponding regression equation for each one.

Spectrophotometric methods manipulating ratio spectra

For RD method, to determine ASP and ROX, carry on the aforementioned procedures to obtain the ratio spectra of the prepared mixtures, then measure the differences in peak amplitudes between selected wavelength pairs (234 nm & 280 nm for ASP and 254 & 297 nm for ROX). The drugs concentrations were calculated using the corresponding regression equation for each one.

While for ^1DD method, to determine ASP and ROX, apply the first derivative steps on the obtained ratio spectra for each mixture using the specified condition. Record measurements at 238.5 nm and 293 nm and calculate the drugs concentrations using the corresponding regression equation for each one, respectively.

Analysis of dosage forms

For ASP tablets

Ten Aspirin protect[®] 100 mg tablets were accurately weighted and finely crushed and homogenously mixed. An amount of powder equivalent to 100 mg of ASP was accurately weighted, transferred into a beaker, 50 mL of methanol was added, then a continuous stirring was applied for 10 min using a magnetic stirrer. The solution was sonicated for 10 min then filtered and accurately transferred into a 100 mL volumetric flask. The volume was completed to the mark by methanol. Additional dilution was carried out to prepare a working standard solution (100 $\mu\text{g/mL}$) that could be used for subsequent preparations.

Different concentration levels within the linearity range were prepared and analyzed using the described procedures of the suggested methods (DW, RD and ^1DD).

For ROX tablets

Ten Xarelto[®] 10 mg tablets were accurately weighted and finely crushed and homogenously mixed. An amount of powder equivalent to 10 mg of ROX was accurately weighted, transferred into a beaker, 50 mL of methanol: DMSO mixture (95:5, v/v) was added, and using a magnetic stirrer the beaker was continuously stirred for 10 min. The solution was sonicated for 10 min then filtered and accurately transferred into a 100 mL volumetric flask, then the volume was completed to the mark by the same solvent.

Different concentration levels were prepared along the linearity range and analyzed using the described procedures of the suggested methods (DW, RD and ^1DD).

For DW, the concentrations of ASP and ROX were calculated by direct substitution of the absorbance difference in their corresponding regression equations. On the other hand, differences in peak amplitude were used for calculations in RD method. Finally, peak amplitudes at the selected wavelengths were substituted in the corresponding regression equations of ^1DD method.

Application to biological fluid (human plasma)

As the two drugs ASP and ROX are co-administered together in a specific treatment regime, we aimed to develop a sensitive and simple method for their simultaneous determination in human plasma.

A volume of 500.0 μL plasma was transferred into two screw capped centrifuge tubes and then separately spiked with 4 mL of ASP and ROX laboratory synthetic mixtures (Intermediate mixtures concentrations: 25: 12.5 $\mu\text{g/mL}$ of ASP: ROX and 75: 37.5 $\mu\text{g/mL}$ of ASP: ROX, respectively). 0.5 mL of methanol as a precipitating agent was added to ensure samples full deproteination. Each sample was vortex mixed for 3 min and centrifuged at 4000 rpm

for 20 min. The supernatants were filtered through a 0.22 μm PVDF filter. Two mL of the filtrate was transferred into a 10-mL volumetric flask and the volume was completed to mark with methanol to obtain mixtures with final concentration of 4: 2 $\mu\text{g}/\text{mL}$ ASP: ROX and 12:6 $\mu\text{g}/\text{mL}$ ASP: ROX, respectively. Blank plasma sample was carried out simultaneously, in an identical manner except for the addition of ASP and ROX mixtures. All the spiked plasma samples were analyzed using the aforementioned procedures. The respective regression equations were then used to determine the drugs concentrations.

Results and discussion

Spectrophotometric methods have proven their superiority and efficiency for quantitative analytical purposes for bi-and/or multi-component mixtures [11, 41–43]. Compared to other analytical procedure such as chromatographic methods; spectrophotometric analysis, can provide easy to apply, accurate, time and cost-effective quantitative analysis procedures for mixtures avoiding chemical pretreatment or sophisticated apparatus criteria, high energy demand, high solvent consumption and influential amount of released hazardous waste [42, 43]. Moreover, spectrophotometric methods can be used to determine the drugs in plasma either directly or after augmentation techniques to reach the maximum blood concentration of a specific analyte [44]. These blessings expand the spectrum of spectrophotometric methods to be used in pharmacokinetics studies and to determine the optimal dose and dosing schedule of a drugs that help achieving the desired therapeutic effect while minimizing the adverse effects.

Variable categorizations can be considered for spectrophotometric methods according to their aims, manipulations, spectral features. Commonly, different spectrophotometric manipulations can be applied for either zero order, normalized or ratio spectra. However, according to multicomponent determinations, mathematical spectrophotometric methods can be classified into five types; signal features enhancement e.g. Derivative; hidden information revealing e.g. Chemometrics; standard addition technique; spiking technique or spectrum addition [41].

To determine ASP and ROX in their novel combination therapy, and to simply address the posed challenge of the significant spectral overlap between ASP and ROX that does not show any chance for clear measurements of neither ASP nor ROX, Fig. 2, it was worthy to find out straightforward approaches that can yield accurate and precise results. Consequently, this work aims to resolve the sever overlapped spectra of ASP and ROX depending on the smart manipulation of the obtained spectra

to determine the selected drugs, simultaneously, in their pure forms, laboratory synthetic mixture and in human plasma samples within their dosage regime ratio. This was achieved by applying three handy, precise, and specific methods with minimum data manipulation and sample preparations. Moreover, statistical studies for the ability of the applied methods in resolving complex spectra was constructed to prove the competency of the suggested method with the official and reported ones and to explore any potential source of invalidity.

The binary mixture was resolved using either zero-order spectra by applying the dual wavelength (DW) method or ratio spectra by applying ratio difference (RD) and derivative ratio (¹DD) methods.

Spectrophotometric methods using zero-order spectra

Dual wavelength method (DW)

The DW method is known for its superb features of being uncomplicated, quick, and precise. It can simply determine the concentration of a particular component in a mixture of other unwanted interfering components. Moreover, the DW method requires minimal data management and consequently, it has a broader range of applications. To utilize this method, the most important requisite is to select a wavelength pair for each drug such that the difference in absorbance is negligible for the interferent and linearly correlated with the target analyte concentrations.

From the overlapping spectra of ASP and ROX, two specific wavelengths were selected for each drug, as shown in Fig. 3. The absorbance difference at 242.5 nm (λ_1) and 255.5 nm (λ_2) was found to be zero for ROX so, its contribution can be repealed and the measurements at these wavelengths are directly proportional to the concentrations of ASP, Fig. 3A. Likewise, the absorbance differences at 252 nm (λ_3) and 286 nm (λ_4) were selected for determination of ROX, where the difference in absorbance at this specific wavelength pair ($A_{286} - A_{252}$) is negligible for ASP, Fig. 3B. The respective regression equation of each drug was obtained by plotting the absorbance difference between either (λ_1, λ_2) or (λ_3, λ_4) against the corresponding concentrations of ASP and ROX, respectively. The calculated regression equations were found to be:

$$\Delta A_{(255.5,242.5)} = 0.0054C + 0.001 \quad r = 0.9998$$

where $\Delta A_{(255.5, 242.5)}$ is the absorbance difference of ASP at 255.5 nm and 242.5 nm, C is the concentration of ASP in $\mu\text{g}/\text{mL}$ and r is the correlation coefficient.

$$\Delta A_{(286,252)} = 0.0291C + 0.0003 \quad r = 0.9997$$

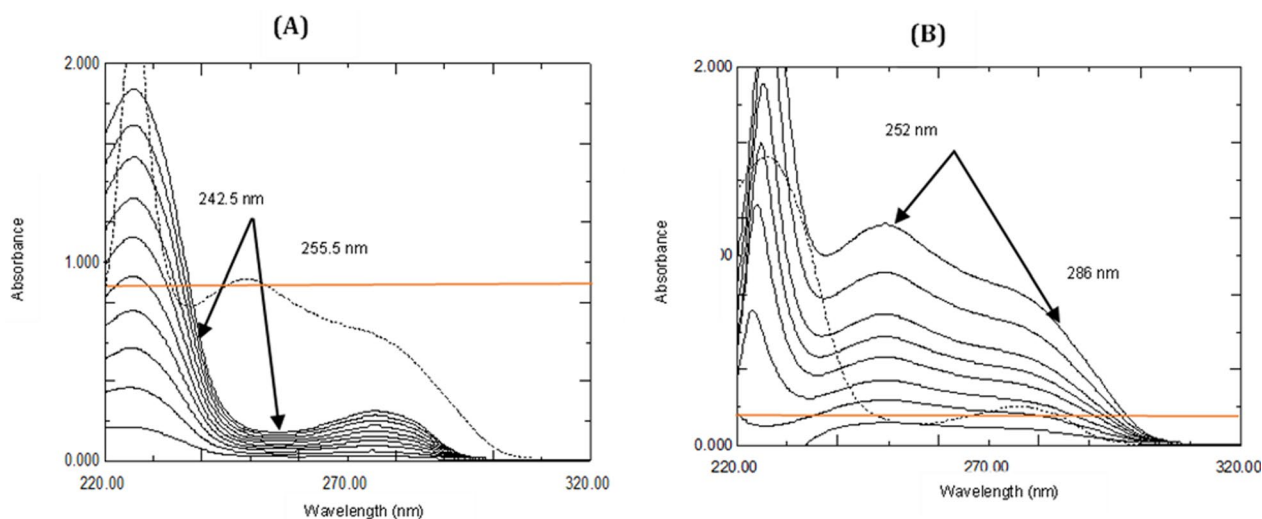


Fig. 3 **A** Zero order absorption spectra of 4–40 $\mu\text{g/mL}$ ASP (—) and 16 $\mu\text{g/mL}$ ROX (....). **B** Zero order absorption spectra of 2–20 $\mu\text{g/mL}$ ROX (—) and 32 $\mu\text{g/mL}$ ASP (....)

where $\Delta A_{(286, 252)}$ is the absorbance difference of ROX at 286 nm and 252 nm, C is the concentration of ROX in $\mu\text{g/mL}$ and r is the correlation coefficient.

The primary downside of DW is its restriction in selecting the wavelength pair for each component that must be limited to those with a constant absorbance of the interfering substance. This necessitates critical measurements; accordingly, any deviation could lead to low sensitivity and accuracy with bad precision. To avoid poor robustness of the method, this drawback has been tackled by applying the ratio difference method that allows constant contribution of the interferent along the whole spectrum.

Spectrophotometric methods manipulating ratio spectra.

Ratio difference method (RD)

In this simple and precise method, the difference between two wavelengths in the ratio spectrum will dispose any interference caused by the component that is used as a divisor [41]. On contrary, RD method allows the determination of a component in its binary mixture at any two wavelengths all over its liner range in the ratio spectrum after using a divisor of the interfering component unlike the DW method that necessitate a constant absorbance of the interfering component at the selected wavelengths.

Upon choosing the divisors, it is crucial to strike a balance between maximum sensitivity and minimal noise. For the analysis of ASP and ROX concentrations in their mixtures, the use of ASP spectrum (36 $\mu\text{g/mL}$) and ROX spectrum (20 $\mu\text{g/mL}$) as a divisor, yielded the most optimum results in terms of average recovery and RSD percentage.

To determine ASP and ROX in their binary mixture, firstly, scan the zero order absorption spectra of the mixtures. For ASP determination, thoughtfully a concentration of ROX spectrum (20 $\mu\text{g/mL}$) was the divisor of choice for the previously scanned spectra. The ratio spectra were produced as shown in Fig. 4A, the amplitudes at 234 nm and 280 nm were measured and subtracted, so the constant contribution of ROX would be cancelled. Then the regression equation used for subsequent determinations of ASP was obtained by plotting the difference in the amplitudes at 234 and 280 nm of the ratio spectra against the corresponding concentrations. The regression equation for ASP was computed and found to be:

$$\Delta P = 0.0221C - 0.0099 \quad r = 0.9998$$

where ΔP is the amplitude difference at the two selected wavelengths, C is the ASP corresponding concentration in $\mu\text{g/mL}$ and r is the correlation coefficient.

Similarly, the ratio spectra for ROX determination were produced using ASP spectrum (36 $\mu\text{g/mL}$) as a divisor and the measurements were recorded at the selected wavelength pair (254 and 297 nm) as shown in Fig. 4B. The regression equation was computed and found to be:

$$\Delta P = 0.1444C + 0.1495r = 0.9999$$

where ΔP is the amplitude difference at the two selected wavelengths, C is the ROX corresponding concentration in $\mu\text{g/mL}$ and r is the correlation coefficient.

Derivative ratio method (.1DD)

The derivative ratio spectra method relies on the step of transformation of the ratio spectrum into its first

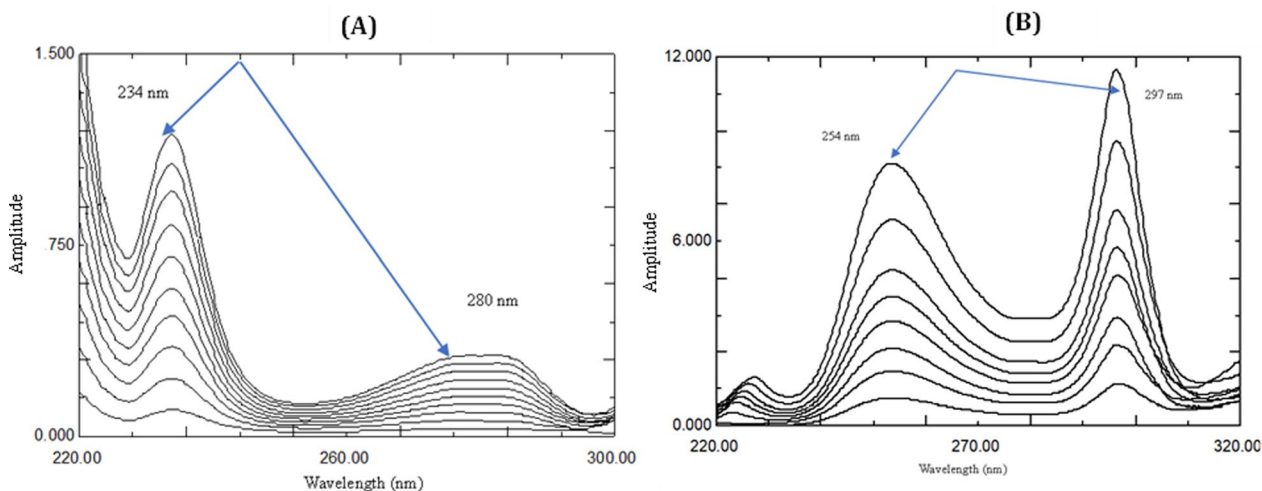


Fig. 4 **A** Ratio spectra of ASP 4–40 µg/mL using 20 µg/mL ROX as a divisor in methanol. **B** Ratio spectra of ROX 2–20 µg/mL using 36 µg/mL ASP as a divisor in methanol

derivative to obtain a spectrum that is independent of the concentration of the interferent. Then measure the amplitude signals of the derivative ratios of the target analyte without any intervention from the other component in the mixture.

For ASP determination, the first derivative of ratio spectra of different concentrations of ASP that obtained using ROX absorption spectrum (20 µg/mL) as a divisor were scanned, then the amplitudes at 238.5 nm were measured that correspond to linear correlation with ASP concentrations as shown in Fig. 5A.

The calibration graph relating the peak amplitudes values at 238.5 nm to the corresponding ASP concentrations was constructed and regression equation was computed and found to be:

$$P_{238.5} = -0.0306C + 0.0277 \quad r = 0.9999$$

where $P_{238.5}$ is the ASP peak amplitudes at 238.5 nm, C is the concentration of ASP in µg/mL and r is the correlation coefficient.

Similarly, for ROX determination, the first derivative of the ratio spectra, obtained using ASP septum (36 µg/mL) as a divisor, were scanned. Several amplitude peaks

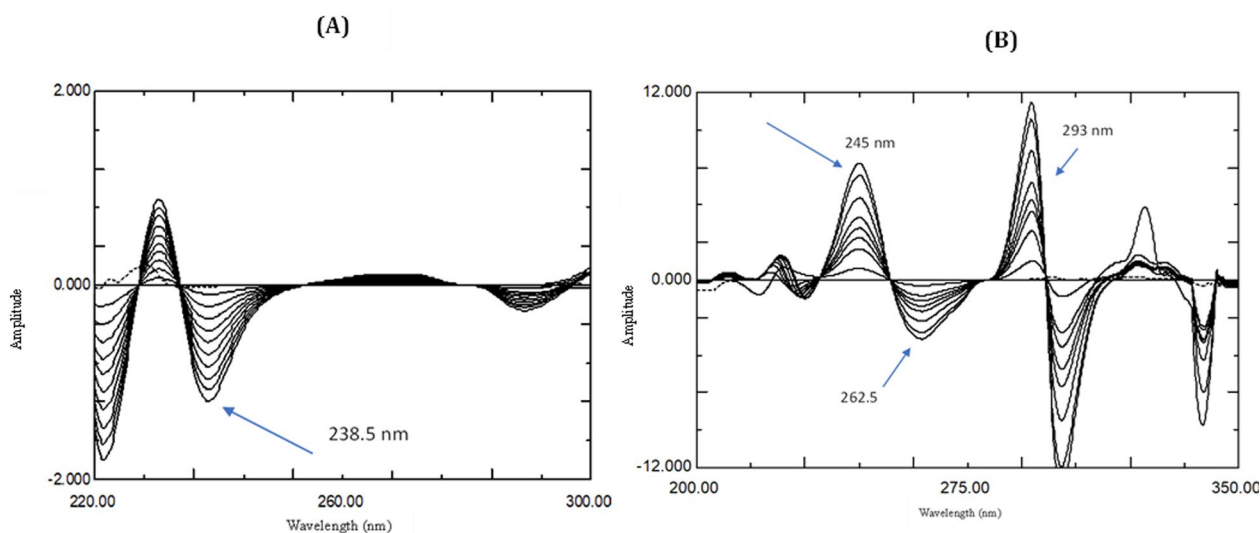


Fig. 5 **A** ¹DD amplitudes of 4–40 µg/mL ASP (—) using 20 µg/mL ROX (....) as a divisor. **B** ¹DD amplitudes of 2–20 µg/mL ROX (—) using 36 µg/mL ASP (....) as a divisor

and troughs of ROX were observed at 245, 262.5 and 293 nm that allow direct determination of ROX without any interference from ASP. All showed good correlation ($r > 0.999$), yet 293 nm was the wavelength of choice considering the fact that it showed the highest sensitivity, as in Fig. 5B.

The calibration graph relating the peak amplitudes values at 293 nm to the corresponding ROX concentrations was constructed and regression equation was computed and found to be:

$$P_{293} = 0.5056C + 0.1667 \quad r = 0.9999$$

where P_{293} is the ROX peak amplitudes at 293 nm, C is the concentration of ROX in $\mu\text{g/mL}$ and r is the correlation coefficient.

For the laboratory synthetic mixtures analysis, zero-order spectra were divided by ROX (20 $\mu\text{g/mL}$) followed by first derivative, and the peak amplitudes measured at 238.5 nm were used to calculate ASP concentrations using the specified regression equation. Then similarly, zero-order spectra were divided by ASP (36 $\mu\text{g/mL}$) spectrum followed by first derivative and peak amplitudes measured at 293 nm were used to calculate ROX concentration using the specified regression equation.

The regression parameters (slope, intercept, and correlation coefficient) were examined to detect the impact of the divisor concentration. Different divisors were tried and ASP (36 $\mu\text{g/mL}$) and ROX (20 $\mu\text{g/mL}$) were found the optimum ones with regard to baseline noise and measurements sensitivity.

The main concept of the chosen spectrophotometric methods is to quantify the target analyte within the studied matrices simply and efficiently without releasing more hazardous chemicals into our biosphere. The most remarkable advantage of DW method is it relies on the intrinsic features of the zero-order spectrum of the analyte. This value makes the determination more reliable, simple and accurate. However, the RD method requires one more graphical manipulation step to obtain the ratio spectra, but it allows more choices for wavelength pairs for the determinations since the interference contribution remains constant. Furthermore, IDD method has a superior merit over the derivative procedure in which it does not need zero crossing or coincidence point for quantitation as well as it creates new sensitive analytical signals (peak or trough) that increase results sensitivity [45]. Additionally, the availability of many maxima and minima provides an extra benefit in detecting the analyte of interest in the presence of other interfering compounds and/or excipients during the assay.

Method validation and statistical analysis

The ICH guidelines for method validation [46] were followed with explicit assumptions of accuracy and precision based on the obtained data. The demonstrated results in Table 1 support the claims of validity of the proposed spectrophotometric methods.

Table 1 Analytical performance data for ASP and ROX determination using the developed spectrophotometric methods

Parameter	ASP			ROX		
	DW	RD	IDD	DW	RD	IDD
λ (nm)	242.5 & 255.5	234 & 280	238.5	252 & 286	254 & 297	293
Range ($\mu\text{g/mL}$)	4-40 $\mu\text{g/mL}$			2-20 $\mu\text{g/mL}$		
Linearity						
Slope	0.0054	0.0221	-0.0306	0.0291	0.1444	0.5056
Intercept	0.001	-0.0099	0.0277	0.0003	0.1495	0.1667
Correlation coefficient	0.9997	0.9997	0.9999	0.9995	0.9998	0.9998
Accuracy ^a						
Mean \pm RSD	100.26 \pm 1.057	100.72 \pm 0.957	99.30 \pm 0.799	99.91 \pm 0.621	101.38 \pm 0.490	99.57 \pm 0.614
Precision (% RSD)						
Repeatability ^b	1.132	1.078	1.213	1.037	1.096	1.158
Intermediate precision ^c	1.288	1.319	1.499	1.395	1.214	1.361
Limits						
Limit of detection, LOD	0.61	0.66	0.53	0.45	0.29	0.26
Limit of quantitation, LOQ	1.85	2	1.63	1.37	0.92	0.79

^a Accuracy was checked using concentrations (10, 14, 22 $\mu\text{g/mL}$) for ASP and (5, 9, 11 $\mu\text{g/mL}$) for ROX

^b and ^c Are the intraday and inter-day respectively ($n = 3$) relative standard deviation of concentrations (4, 16, 24 $\mu\text{g/mL}$) for ASP and (4, 8, 12 $\mu\text{g/mL}$) for ROX

Analytical application of the proposed spectrophotometric methods for dosage forms

The proposed methods were successfully implemented to determine ASP and ROX in their commercial tablets. Application of standard addition technique was done to confirm the accuracy of the three methods as stated in Table 2.

Analytical application of the proposed spectrophotometric methods to spiked human plasma

The proposed methods were effectively applied for the determination of the novel combination therapy of ASP and ROX in human plasma. Plasma samples typically require deproteinization to eliminate matrix effect. We used a simple liquid/liquid extraction step which was very suitable for our suggested methods as the standards, and the binary mixtures are dissolved in the organic solvent that will help the deprotonation of the plasma without any additional reagents or procedures.

Standard addition technique was applied for the study by spiking suitable increments of each drug to blank plasma. This technique is more advisable for drugs with low plasma concentration levels since it augments the drugs' concentrations to reach their C_{max} [47, 48] and facilitates their quantitative analysis in human fluids. The matrix effect was assessed by comparing the absorption spectrum of blank plasma and the obtained

absorption spectra of the prepared mixtures of ASP & ROX either in plasma or in methanol in the spiked concentrations (zero order absorption spectra, the ratio spectra after division by ROX 20 $\mu\text{g}/\text{mL}$ spectrum or 36 $\mu\text{g}/\text{mL}$ ASP spectrum and the first derivative ratio spectra), Fig. 6A–C. All these spectra proved the applicability of the proposed methods for the determination of the selected drugs in the biological matrix of plasma without any interference at the selected wavelengths for measurements. Furthermore, Table 2 evinces the mean recoveries and RSD% of both drugs that support the methods' credibility.

Statistical analysis

The obtained results of the suggested methods for the determination of ASP and ROX were sufficiently good. Nonetheless, statistical evaluation of these results remains crucial, as it offers insight into whether the methods can be applied in QC & bioequivalent analysis. Statistical comparison with results obtained by applying the USP official method for ASP [40] and the reported HPLC method [34] was performed as shown in Table 3. Regarding *t*- and *F*-values, the null hypothesis was accepted, and the proposed methods showed good accuracy and excellent precision with no significant interference from pharmaceutical or biological matrices.

Table 2 Determination of ASP and ROX in laboratory-prepared mixtures, marketable sample, human plasma and application of standard addition technique

Sample	Method		
	DW	RD	1DD
ASP			
Laboratory prepared mixtures (n=5)*	100.12 ± 0.871	99.93 ± 0.854	99.77 ± 1.221
Aspirin protect 100 (BTAB235)	99.21 ± 0.699	99.79 ± 1.042	99.98 ± 1.525
Standard addition of dosage form	98.08 ± 0.943	99.73 ± 1.644	100.82 ± 0.650
Human Plasma ^{a,**}			
4 $\mu\text{g}/\text{mL}$	99.49 ± 0.805	100.87 ± 0.515	99.81 ± 1.214
12 $\mu\text{g}/\text{mL}$	99.16 ± 0.953	98.85 ± 1.146	100.44 ± 0.589
Standard addition of plasma ^{a,**}	101.01 ± 0.484	100.04 ± 1.111	99.42 ± 0.921
ROX			
Laboratory prepared mixtures (n=5)*	99.88 ± 1.077	99.93 ± 0.854	99.77 ± 1.221
Xarelto 10 mg (NDC 50458-580-30)	100.74 ± 0.952	100.50 ± 1.013	100.80 ± 0.437
Standard addition of dosage form	100.67 ± 1.363	99.95 ± 1.040	98.96 ± 0.491
Human Plasma ^{a,**}			
2 $\mu\text{g}/\text{mL}$	100.24 ± 1.002	99.59 ± 0.892	100.38 ± 0.911
4 $\mu\text{g}/\text{mL}$	100.05 ± 1.071	100.91 ± 0.819	101.06 ± 0.794
Standard addition for plasma ^{a,**}	99.92 ± 1.447	100.12 ± 1.589	100.30 ± 1.033

* 5 sets for each method, each of 3 replicates

** Mean ± SD %

^a Each result is the average of three separate determinations

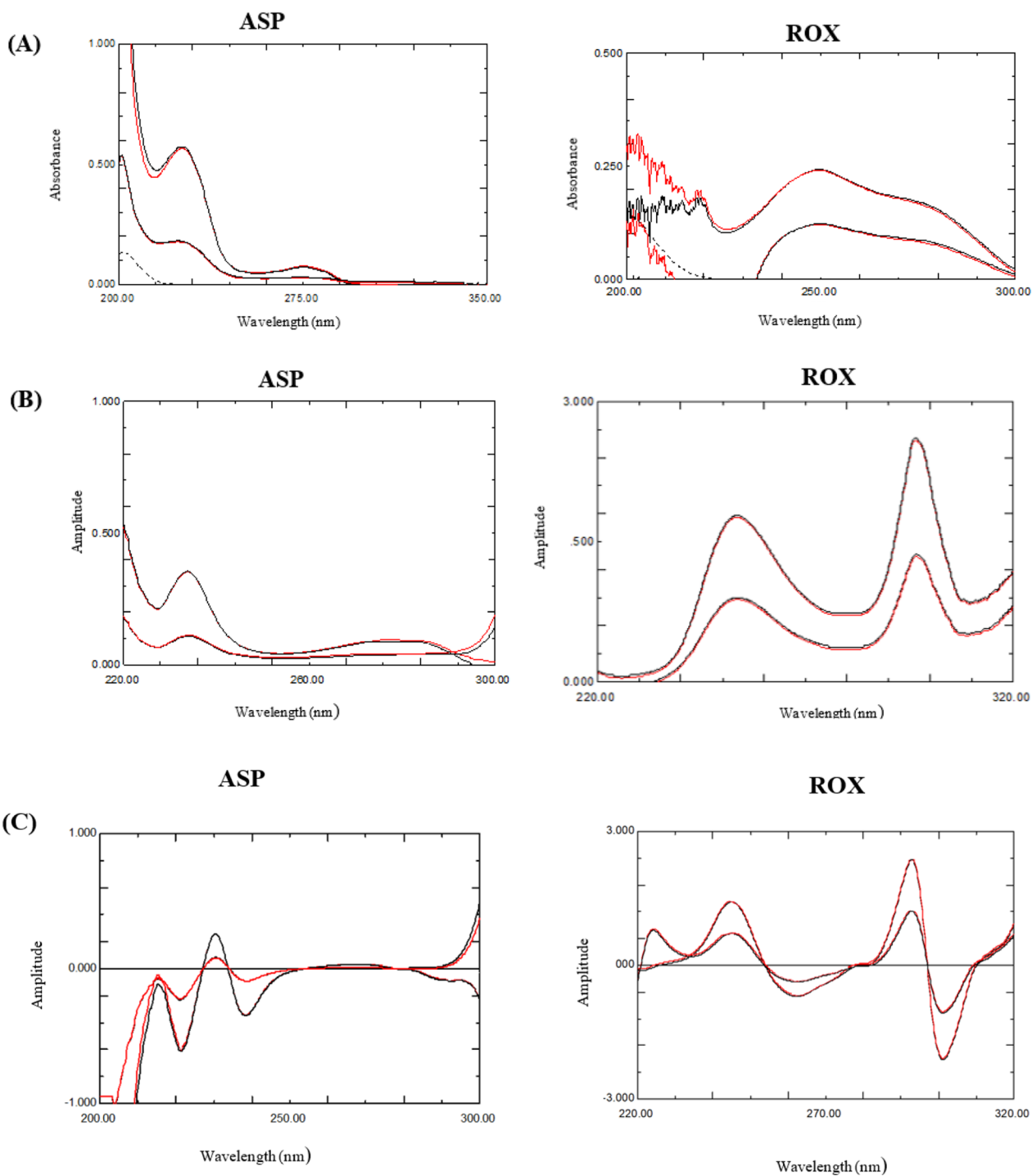


Fig. 6 **A** Zero order absorption spectra of Blank plasma (...), 4 and 12 $\mu\text{g}/\text{mL}$ ASP in methanol (black) and in plasma (red). And 2 and 4 $\mu\text{g}/\text{mL}$ ROX in methanol (black) and in plasma (red). **B** Ratio spectra of 4 and 12 $\mu\text{g}/\text{mL}$ ASP in methanol (black) and plasma (red) using ROX 20 $\mu\text{g}/\text{mL}$ as divisor. And 2 and 4 $\mu\text{g}/\text{mL}$ ROX in methanol (black) and plasma (red) using ASP 36 $\mu\text{g}/\text{mL}$ as divisor. **C** ^1DD of 4 and 12 $\mu\text{g}/\text{mL}$ ASP in methanol (black) and plasma (red) using ROX 20 $\mu\text{g}/\text{mL}$ and ^1DD of 2 and 4 $\mu\text{g}/\text{mL}$ ROX in methanol (black) and plasma (red) using ASP 36 $\mu\text{g}/\text{mL}$ as divisor.

Methods comparative study

The applied spectrophotometric methods offer competitor analytical tools for the determination of such a new

mixture. Despite some limitations for each method, they are considered the superior choice in QC labs, bioavailability labs, Table 4.

Table 3 Statistical analysis of the results obtained by the developed spectrophotometric methods and official and reported methods for determination of ASP and ROX in their pure powdered form

Parameter	ASP				ROX				
	DW	RD	1DD	Official ^a	DW	RD	1DD	Reported ^b	
Mean	100.26	100.11	99.7	100.22	100.06	100.24	99.67	100.48	
SD	1.00	1.05	0.89	0.95	0.97	0.68	0.706	0.92	
n	10	10	10	6	8	8	8	6	
Variance	1.00	1.11	0.79	0.90	0.94	0.47	0.49	0.84	
Student's t-test	0.078 (2.145)*	0.205 (2.145)*	1.215 (2.145)*		0.855 (2.179)*	0.562 (2.179)*	1.888 (2.179)*		
F	1.10 (4.77)*	1.23 (4.77)*	1.14 (4.77)*		1.11 (4.88)*	1.79 (4.88)*	3.81 (4.88)*		

* The figures in parenthesis are the corresponding theoretical values at P=0.05

^a Official titrimetric USP method

^b Reported HPLC method using C18 column isocratic elution using ACN: H₂O (55:45 V/V), UV detection at 249 nm

Table 4 Advantages and outcomes of the proposed spectrophotometric methods

Method	Advantages & outcomes	Limitations
DW	<ul style="list-style-type: none"> • Can determine the co-administered ASP &ROX successfully in different matrices • Applied on zero order spectra, consequently less noise is obtained • No need for further manipulation so, it does not require special software • Has the least mathematical slope & intercept of linear equation, so low interference and error index are assured • Has RSD% less than 2 which offers accurate and precise determinations 	<ul style="list-style-type: none"> • Applied on Zero order spectra, so some minor features of the spectra may be neglected • The wavelength pairs of choice may be critical which propose meticulous analysis
RD	<ul style="list-style-type: none"> • Can determine the co-administered ASP &ROX successfully in different matrices • Applied on ratio spectra, that keeps the potential interference constant throughout the whole spectrum • Allow wide choice of wavelength pairs for determination • Critical determinations are avoided • Has low mathematical slope & intercept of linear equation, so low interference and error index are obtained • Has RSD% less than 2 which offers accurate and precise determinations 	<ul style="list-style-type: none"> • Ratio spectra may enhance base line noise • It needs one more manipulation step • It requires divisor selection optimization
1DD	<ul style="list-style-type: none"> • Can determine the co-administered ASP &ROX successfully in different matrices • Applied on ratio spectra, that keeps the potential interference constant throughout the whole spectrum • Derivatization cancels the interfering contribution throughout the spectra • Derivative manipulation may enhance some minor features that permit clear determination • Allow variable wavelengths for quantitation • Has low mathematical slope & intercept of linear equation, so low interference and error index are obtained • Creates new sensitive analytical signals giving the chance of more sensitive results • Has RSD% less than 2 which offers accurate and precise determinations 	<ul style="list-style-type: none"> • Ratio spectra with derivative steps may enhance base line noise • It needs more manipulation steps • It requires divisor and derivatization parameters optimization

Ecological consideration and greenness assessment

During the planning stage of the analytical methodology, great attention was given to evaluate its ecological impact [49]. Figure 7A elaborates the selection factors considered for the suggested method within the ecological paradigm framework. We chose a direct spectrophotometric method, as it offered a significant advantage of eliminating cumbersome sample preparation and separation

steps, resulting in reduced use of hazardous solvents compared to Liquid chromatographic methods [15, 50].

For the solvent selection, it was the most crucial step. Rivaroxaban solubility is challenging as it is water insoluble and sparingly soluble in alcohols like ethanol and methanol. This was overcome by use of co-solvents to improve solubility without excessive use of the less green solvents. After several solubility trials, we have selected to use methanol with minimum volume of

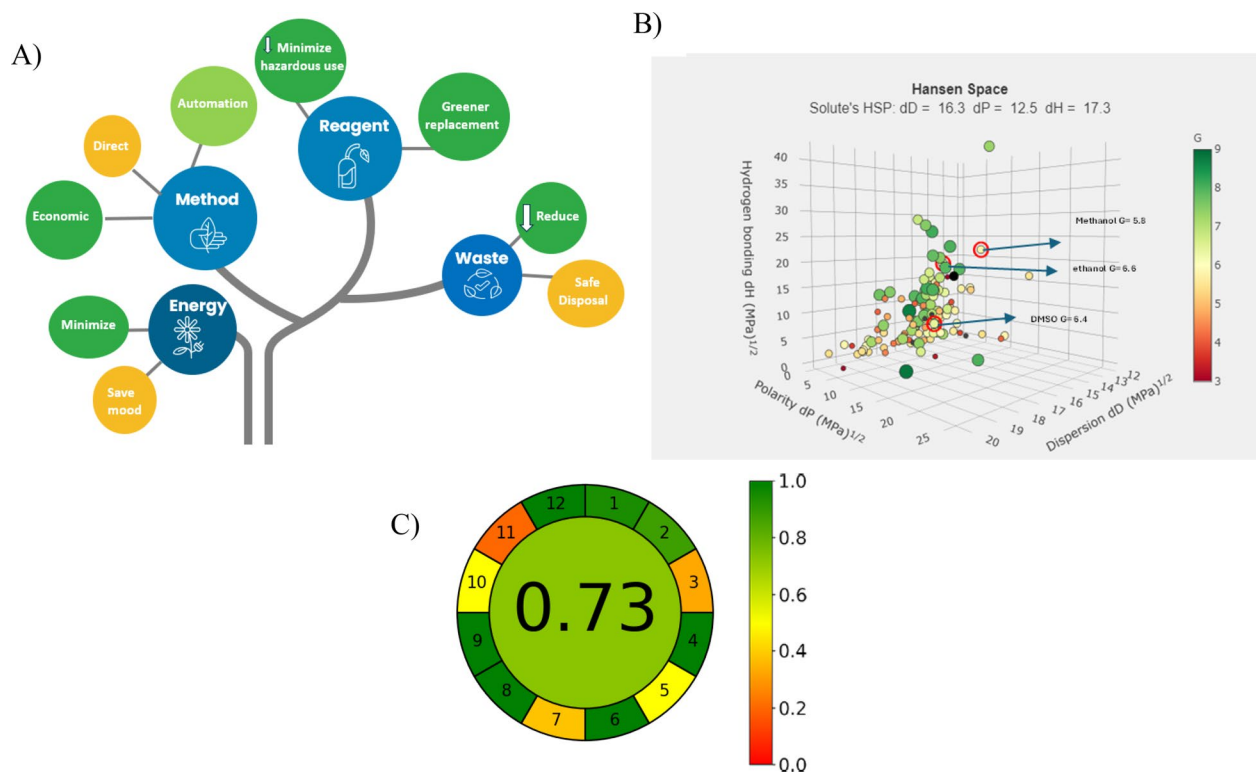


Fig. 7 **A** Contemplating selection factors for the suggested method within the ecological paradigm framework. **B** G scores by GSST for ethanol, methanol and DMSO. **C** The Analytical Greenness Calculator (AGREE) for Greenness assessment

DMSO that guarantee a full solubility of the drug even after storage proved by intermediate precisions and stability checking of the stock solution. This practice echoed the recommendation of rivaroxaban solubility reported in [51] that recommends the binary solvents mixture of DMSO and Methanol. We used the greenness score from the GSK Solvent Sustainability Guidelines [52], which presents a numerical evaluation of solvents through a Composite Score reflecting various criteria (G) which calculated using a free calculator <http://green-solvent-tool.herokuapp.com/> according to the equation $G = \sqrt[4]{(H \times S \times E \times W)}$. Ethanol scored $G = 6.6$ while DMSO 6.4 and Methanol 5.8 (Fig. 7B). However, the use of ethanol, a more environmentally friendly option but was not conducive to the drug's solubility. This necessitates a balance between adhering to eco-friendly practices and ensuring the method's practicality. In addition, the use of methanol facilitates the plasma extraction procedure [53] as it acts as the precipitating agent for the proteins without the need of various reagents with various environmental impacts. Any interference from the used solvent can simply be removed using a blank.

In terms of energy use, being spectrophotometric methods, we consumed minimal energy due to quick sample measurements and the offline spectra analysis and manipulation. Moreover, for the produced waste, compared to alternative chromatographic techniques, it generated significantly less waste, making safe waste disposal easier to safeguard.

To assess the greenness profile, we have used the latest developed tool, the Analytical Greenness calculator (AGREE) [54]. It utilizes a simple algorithm that uses a scale from 0 to 1 to represent the method's overall environmental sustainability, along with a pictorial representation directly linked to the 12 GAC rules. The automatically generated pictogram is composed of twelve sections, each featuring a unique color gradient that spans from dark green (value=1) to dark red (value=0). At the center of the pictogram, the total score is displayed as a fraction, with values ranging from zero to one indicating how close to the ideal green value (value=1). The AGREE software delivered a detailed assessment of the entire analytical process associated with each principle of green chemistry and the calculation is automated using free software

(<https://mostwiedzy.pl/AGREE>), [54]. It emphasized the most vulnerable segments of the analytical procedures, pinpointing where further adjustments are necessary to enhance sustainability. Our suggested spectrophotometric methods proved its agreement with the GAC principles through a comprehensive assessment of its efficiency, practicality, and environment friendliness. The Agree score is 0.73, the missing points are mainly related to miniaturization and the reagents which cannot be fully avoided especially with the solubility issues of the studied drug. The score still ensures an overall minimal ecological footprint of the suggested methods as shown in Fig. 7C.

Concluding remarks

The recommended methods are considered the first spectrophotometric methods to determine this novel combination therapy of aspirin and rivaroxaban. They are selective, accurate, sensitive, and more appropriate, easier, and cheaper for determination of the studied mixture when compared to any chromatographic method. Even though the C_{max} does not lie within the working concentration range of the suggested methods, applying the spiking technique allows their accurate analysis. The proposed methods provide a direct, cost effective and simple alternative for determination of the combination therapy without preliminary separation or elaborate treatment associated with chromatographic methods.

Linearity of the calibration graphs and adherence to Beer's law were confirmed by the small intercepts and the high values of correlation coefficients. The three methods demonstrated reliability and accuracy, with minimum data manipulation requirement. The DW method was the simplest, while the DR method has broader application as it does not require constant absorbance of the interfering components at the selected wavelengths. The suggested methods were successfully applied to the analysis of ASP and ROX in spiked human plasma.

Author contributions

HMM: Conceptualization, methodology, formal analysis, validation, data curation, writing—original draft, writing—review & editing, visualization. HME: Conceptualization, methodology, validation, formal analysis, data curation, writing—original draft, writing—review & editing, visualization.

Funding

Open access funding provided by The Science, Technology & Innovation Funding Authority (STDF) in cooperation with The Egyptian Knowledge Bank (EKB).

Availability of data and materials

Any supplementary data will be made available on request.

Declarations

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare no conflict of interest.

Received: 5 October 2023 Accepted: 8 February 2024

Published online: 19 February 2024

References

- Lecouturier J, Scott J, Rousseau N, Stansby G, Sims A, Allen J. Peripheral arterial disease diagnosis and management in primary care: a qualitative study. *BJGP Open*. 2019. <https://doi.org/10.3399/bjgpopen19X101659>.
- Fowkes FGR, Rudan D, Rudan I. Comparison of global estimates of prevalence and risk factors for peripheral artery disease in 2000 and 2010: a systematic review and analysis. *Lancet*. 2013;382(9901):1329–40. [https://doi.org/10.1016/S0140-6736\(13\)61249-0](https://doi.org/10.1016/S0140-6736(13)61249-0).
- Criqui MH, Aboyans V. Epidemiology of peripheral artery disease. *Circ Res*. 2015;116(9):1509–26. <https://doi.org/10.1161/CIRCRESAHA.116.303849>.
- Anand SS, Hiatt W, Dyal L, Bauersachs R, Berkowitz SD, Branch KRH, Debus S, Fox KAA, Liang Y, Muehlhofer E, Nehler M, Haskell LP, Patel M, Szarek M, Yusuf S, Eikelboom J, Bonaca MP. Low-dose rivaroxaban and aspirin among patients with peripheral artery disease: a meta-analysis of the COMPASS and VOYAGER trials. *Eur J Prev Cardiol*. 2022;29(5):e181–9. <https://doi.org/10.1093/eurjpc/zwab128>.
- FDA Approves Expanded Peripheral Artery Disease (PAD) Indication for XARELTO® (rivaroxaban) Plus Aspirin to Include Patients After Lower-Extremity Revascularization (LER) Due to Symptomatic PAD. <https://www.fda.gov/news-center>. Accessed 2 Apr 2023.
- Hamzah K, et al. Low-dose aspirin and rivaroxaban combination therapy to overcome aspirin non-sensitivity in patients with vascular disease. *Front Cardiovasc Med*. 2022;9: 912114. <https://doi.org/10.3389/fcvm.2022.912114>.
- Senzana S, Gordana Z, Aleksandra N, Senzana B, Salvinca M. Quantitative analysis of acetylsalicylic acid in commercial pharmaceutical formulations and Human control serum using kinetic spectrophotometry. *Acta chem solv*. 2008;55:508–15.
- Schrör K. Aspirin and platelets: the antiplatelet action of aspirin and its role in thrombosis treatment and prophylaxis. *Semin Thromb Hemost*. 2007;199723(4):349–56. <https://doi.org/10.1055/s-2007-996108>.
- Perzborn E, Roehrig S, Straub A, Kubitzka D, Mueck W, Laux V. Rivaroxaban: a new oral factor Xa inhibitor. *Arterioscler Thromb Vasc Biol*. 2010;30:376–81.
- Zhang Q, Ding Q, Yan S. Fatal adverse events of rivaroxaban combined with aspirin: an analysis using data from Vigibase. *Eur J Clin Pharmacol*. 2022;78:1521–6. <https://doi.org/10.1007/s00228-022-03357-4>.
- Murtaza G, Khan SA, Shabbir A, Mahmood A, Asad MHHB, Farzana K, Hussain I. Development of a UV-spectrophotometric method for the simultaneous determination of aspirin and paracetamol in tablets. *Scientific research and Essays*. 2011;6(2):417–21.
- Moş AC, Soponar F, Medvedovici A, Sârbu C. Simultaneous spectrophotometric determination of aspirin, paracetamol, caffeine, and chlorphenamine from pharmaceutical formulations using multivariate regression methods. *Anal Lett*. 2010;43(5):804–13.
- Elmasry MS, Hassan WS, El-Mammli MY, Badrawy M. Earth friendly spectrophotometric methods based on different manipulation approaches for simultaneous determination of aspirin and omeprazole in binary

- mixture and pharmaceutical dosage form: comparative statistical study. *Spectrochim Acta Part A Mol Biomol Spectrosc.* 2022;266: 120436.
14. Motan G, Puia A. Studies of different types of aspirin by spectrophotometric methods. *Acta Chemica Iasi.* 2014;22(2):155–64.
 15. Kayali Z, Obaydo RH, Alhaj SA. Spider diagram and sustainability evaluation of UV-methods strategy for quantification of aspirin and sildenafil citrate in the presence of salicylic acid in their bulk and formulation. *Heliyon.* 2023;9(4): e15260. <https://doi.org/10.1016/j.heliyon.2023.e15260>.
 16. Gujarathi SC, Shah AR, Jagdale SC, Datar PA, Choudhari VP, Kuchekar BS. Spectrophotometric simultaneous determination of aspirin and Ticlopidine in combined tablet dosage form by first order derivative spectroscopy, area under curve (AUC) and ratio derivative spectrophotometric methods. *Int J Pharm Sci Rev Res.* 2010;3(1):115–9.
 17. Mullangi R, Sharma S, Srinivas NR. Review of HPLC methods and HPLC methods with mass spectrometric detection for direct determination of aspirin with its metabolite (s) in various biological matrices. *Biomed Chromatogr.* 2012;26(8):906–41.
 18. Patel SM, Patel CN, Patel VB. Stability-indicating HPLC method for simultaneous determination of aspirin and prasugrel. *Indian J Pharm Sci.* 2013;75(4):413–9. <https://doi.org/10.4103/0250-474X.119816>.
 19. Kalmár É, Gyuricza A, Kunos-Tóth E, Szakonyi G, Dombi G. Simultaneous quantification of paracetamol, acetylsalicylic acid and papaverine with a validated HPLC method. *J Chromatogr Sci.* 2014;52(10):1198–203. <https://doi.org/10.1093/chromsci/bmt177>.
 20. Franeta JT, Agbaba D, Eric S, Pavkov S, Aleksic M, Vladimirov S. HPLC assay of acetylsalicylic acid, paracetamol, caffeine and phenobarbital in tablets. *Farmaco.* 2002;57(9):709–13. [https://doi.org/10.1016/s0014-827x\(02\)01265-x](https://doi.org/10.1016/s0014-827x(02)01265-x).
 21. Londhe SV, Deshmukh RS, Mulgund SV, Jain KS. Development and validation of a reversed-phase HPLC method for simultaneous determination of aspirin, atorvastatin calcium and clopidogrel bisulphate in capsules. *Indian J Pharm Sci.* 2011;73(1):23.
 22. Rajavel R, Ganesh M, Jagadeeswaran M, Srinivasan K, Valarmathi J, Sivakumar T. RP-HPLC method for the simultaneous determination of aspirin, atorvastatin and pioglitazone in capsule dosage form. *Asian J Res Chem.* 2008;1(1):40–2.
 23. Prakash K, Kalakuntla RR, Sama JR. Rapid and simultaneous determination of aspirin and dipyridamole in pharmaceutical formulations by reversed-phase high performance liquid chromatography (RP-HPLC) method. *Afr J Pharm Pharmacol.* 2011;5(2):244–51.
 24. Elmasry MS, Blagbrough IS, Rowan MG, Saleh HM, Kheir AA, Rogers PJ. Quantitative HPLC analysis of mebeverine, mesalazine, sulphasalazine and dispersible aspirin stored in a Venalink monitored dosage system with co-prescribed medicines. *J pharma biomed anal.* 2011;54(4):646–52.
 25. Abdelaleem EA, Abou El Ella DA, Mahmoud AM, Abdelhamid NS. Green analysis of newly approved binary omeprazole/aspirin mixture in presence of aspirin impurity using ultra-high-performance liquid chromatography and thin-layer chromatography methods. *Biomed Chromatogr.* 2021;35(2): e4986.
 26. Kashid AM, Kolhe OH. Simultaneous densitometric determination of Aspirin and Omeprazole by high-performance thin-layer chromatography. *JPC J Planar Chromatogr Modern TLC.* 2019;32:501–4.
 27. Shahin M. Application of TLC densitometric method for simultaneous determination of aspirin and omeprazole in pharmaceutical preparation. *Innoriginal Int J Sci.* 2018;5:47–50.
 28. Ghadimi H, Tehrani RM, Basirun WJ, Ab Aziz NJ, Mohamed N, Ab GS. Electrochemical determination of aspirin and caffeine at MWCNTs-poly-4-vinylpyridine composite modified electrode. *J Taiwan Inst Chem Eng.* 2016;65:101–9.
 29. Sartori ER, Medeiros RA, Rocha-Filho RC, Fatibello-Filho O. Square-wave voltammetric determination of acetylsalicylic acid in pharmaceutical formulations using a boron-doped diamond electrode without the need of previous alkaline hydrolysis step. *J Braz Chem Soc.* 2009;20(2):360–6. <https://doi.org/10.1590/S0103-50532009000200022>.
 30. Celebier M, Kaynak SN, Altnoz S, Sahin S. UV spectrophotometric method for determination of the dissolution profile of rivaroxaban. *Dissolution Technol.* 2014;21:56–9.
 31. Sekaran CB, Bind VH, Damayanthi MR, Sireesha A. Development and validation of UV spectrophotometric method for the determination of rivaroxaban. *Der Pharma Chem.* 2013;5:1–5.
 32. Prawez A, Essam E, Muzaffar I, Khalid MD, Mostafa GAE, Alqarni MH, Foudah AI, Shakeel F. Ecofriendly densitometric RP-HPLC method for determination of rivaroxaban in nanoparticle formulations using green solvents. *RSC Adv.* 2020;10:2133–40. <https://doi.org/10.1039/C9RA07825H>.
 33. Lories IB, Mostafa AA, Girges MA. High performance liquid chromatography, TLC densitometry, first derivative and first derivatantsolvents. o spectrophotometry for determination of rivaroxaban and its alkaline degradates in bulk powder and its tablets. *J Chromatogr Sep Tech.* 2013;4:E202.
 34. Mustafa C, Tuba R, Engin K, Sacide A. HPLC method development and validation for estimation of rivaroxaban in pharmaceutical dosage forms. *Braz J Pharm Sci.* 2013;49:359–66.
 35. Chandra K, Satya P, Dhana A, Anupama C, Devanaboyina N. A new method for development and validation for analysis of rivaroxaban in formulation by RPHPLC. *Res Desk.* 2012;1:24–33.
 36. Kasad PA, Muralikrishna KS. Method development and acid degradation study of rivaroxaban by RP-HPLC in bulk. *Asian J Pharm Anal.* 2013;3:62–5.
 37. Hadagali MD. Determination of rivaroxaban in pure, pharmaceutical formulations and human plasma samples by RP-HPLC. *Int J Adv Pharm Anal.* 2015;5:65–8.
 38. Rohde G. Determination of rivaroxaban-a novel, oral, direct Factor Xa inhibitor in human plasma by high-performance liquid chromatography-tandem mass spectrometry. *J Chromatogr B.* 2008;872:43–50.
 39. Electro I, Celebier MS, Altnoz S. Electrochemical behaviour investigation and square-wave voltammetric determination of rivaroxaban in pharmaceutical dosage forms. *Anal Methods.* 2014;6:9397–403.
 40. The United States Pharmacopeia and The National Formulary. U.S.; Pharmacopeial Convention: 2019. Rockville MD, USA: USP 42-NF 37.
 41. Obaydo RH, Al Zakri DJ, Sakur AA, et al. Ultraviolet spectrophotometric methods for the determination of the minor component presented in fixed-dose pharmaceutical combinations through the last two decades (2000–2020). *Futur J Pharm Sci.* 2021;7:44. <https://doi.org/10.1186/s43094-021-00192-9>.
 42. Yehia AM, Mohamed HM. Chemometrics resolution and quantification power evaluation: application on pharmaceutical quaternary mixture of paracetamol, guaifenesin, phenylephrine and p-aminophenol. *Spectrochim Acta Part A Mol Biomol Spectrosc.* 2016;152:491–500.
 43. Mohamed HM, Lamie NT. Application and validation of superior spectrophotometric methods for simultaneous determination of ternary mixture used for hypertension management. *Spectrochim Acta Part A Mol Biomol Spectrosc.* 2016;155:103–10. <https://doi.org/10.1016/j.saa.2015.11.001>.
 44. Redasani VK, Patel PR, Marathe DY, Chaudhari SR, Shirkhedkar AA, Surana SJ. A review on derivative UV-spectrophotometry analysis of drugs in pharmaceutical formulations and biological samples review. *J Chil Chem Soc.* 2018;63(3):4126–34. <https://doi.org/10.4067/s0717-97072018000304126>.
 45. Lotfy HM, Obaydo RH, Mohamed EH. Environmentally sustainable computationally spectrophotometric resolution strategy for analysis single-tablet regimens of antihypertension with overlapped spectra. *Talanta Open.* 2023;7: 100226.
 46. ICH Harmonized Tripartite Guideline. Validation of analytical procedures: text and methodology Q2. R1; 2005
 47. Nagelschmitz J, Blunck M, Kraetzschmar J, Ludwig M, Wensing G, Hohlfeld T. Pharmacokinetics and pharmacodynamics of acetylsalicylic acid after intravenous and oral administration to healthy volunteers. *Clin Pharmacol Adv App.* 2014;6:51–9. <https://doi.org/10.2147/CPAA.S47895>.
 48. Mueck W, Stampfuss J, Kubitzka D, Becka M. Clinical pharmacokinetic and pharmacodynamic profile of rivaroxaban. *Clin Pharmacokinet.* 2014;53(1):1–16. <https://doi.org/10.1007/s40262-013-0100-7>.
 49. Mohamed HM. Green, environment-friendly, analytical tools give insights in pharmaceuticals and cosmetics analysis. *TrAC, Trends Anal Chem.* 2015;66:176–219.
 50. Plotka-Wasylika J, Mohamed HM, Kurowska-Susdorf A, Dewani R, Fares MY, Andrich V. Green analytical chemistry as an integral part of sustainable education development. *Curr Opin Green Sust Chem.* 2021;31: 100508. <https://doi.org/10.1016/j.cogsc.2021.100508>.
 51. Jeong J, Ha E, Park H, Lee S, Kim M. Measurement and correlation of solubility of rivaroxaban in dichloromethane and primary alcohol binary solvent mixtures at different temperatures. *J Mol Liq.* 2022;357: 119064.

52. Larsen C, Lundberg P, Tang S, Rafols-Ribe J, Sandstrom A, Mattias Lindh E, Wang J, Edman L. A tool for identifying green solvents for printed electronics. *Nat Commun.* 2021;12:4510. <https://doi.org/10.1038/s41467-021-24761-x>.
53. Kong R. 17—LC/MS application in high-throughput ADME screen. *Sep Sci Technol.* 2005;6:413–46.
54. Pena-Pereira P, Wojnowski W, Tobiszewski M. AGREE-analytical GREEnness metric approach and software. *Anal Chem.* 2020;92:10076–82. <https://doi.org/10.1021/acs.analchem.0c01887>.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.